

# 1,1'-(1,32-Dotriacontanediyl)bis[2-acetyl-*sn*-glycero(3)phosphocholine]: A Long Persisting Agonist as a Potential Antihypertension Agent

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(Received October 28, 1988)

**Synopsis.** A dimer of platelet activating factor (PAF), 1,1'-(1,32-dotriacontanediyl)bis[2-acetyl-*sn*-glycero(3)phosphocholine] (**7**), was synthesized and examined for platelet aggregation and antihypertension activities. The compound **7** showed both activities with the maximum levels of 1/50–1/100-fold of those of PAF, but the antihypertension effect of **7** persisted much longer and appeared with time lag of 1–3 min. The unique expression of the activities was discussed in conjunction with the molecular structure.

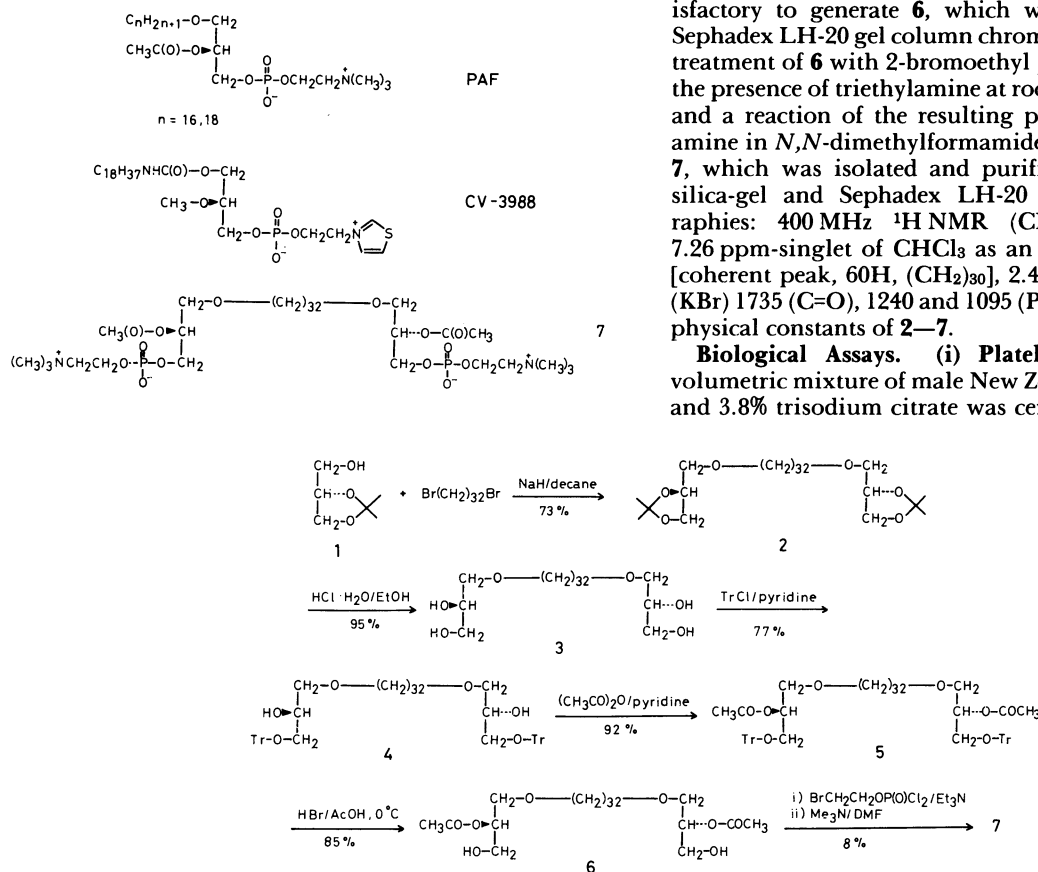
1-Alkyl-2-acetyl-*sn*-glycero(3)phosphocholine (alkyl:  $C_nH_{2n+1}$ ,  $n=16$  and 18) is released from IgE-sensitized basophiles. The lipids are potent platelet activating factor (PAF)<sup>1,2</sup> and serve as chemical mediators in physiological processes such as anaphylaxis and inflammation.<sup>3</sup> Various agonists have been synthesized to reveal the following structural features for the biological activity;<sup>4</sup> (a) a long chain-alkyl ether linkage at the *sn*-1-O atom,<sup>5,6</sup> (b) short chain acylation ( $C_{2-4}$ ) at the *sn*-2-O atom,<sup>7,8</sup> and (c) a

quaternary ammonium residue as small as  $(CH_3)_3N^+$  of the phosphorylcholine moiety at the *sn*-3 position.<sup>9,10</sup> In this paper we wish to report preparation and biological activities (platelet activation and antihypertension) of a new agonist of PAF, 1,1'-(1,32-dotriacontanediyl)bis[2-acetyl-*sn*-glycero(3)phosphocholine] (**7**). The bis(glycerophospholipid) may be considered as a dimeric version of PAF.

## Experimental

**Synthesis.** 2,3-Isopropylidene-*sn*-glycerol (**1**), after conversion into the sodium alkoxide, was heated with 1,32-dibromodotriacontane in decane at 140 °C. The resulting **2** was transformed via **3** and **4** into **5** by a series of usual reactions including acid catalyzed deisopropylidenation, tritylation and acetylation in an overall yield of 73%. Removal of the trityl groups of **5** required considerable experimentation since the acetyl group at the *sn*-2-O position tended to rearrange into the *sn*-3-O position.<sup>11</sup> A treatment of a benzene–ethyl acetate solution of **5** with 25% hydrogen bromide–acetic acid at 0 °C for 50 s was satisfactory to generate **6**, which was isolated by means of Sephadex LH-20 gel column chromatography. A subsequent treatment of **6** with 2-bromoethyl phosphorodichloridate in the presence of triethylamine at room temperature overnight and a reaction of the resulting phosphate with trimethylamine in *N,N*-dimethylformamide at 55 °C for 4 d provided **7**, which was isolated and purified by a combination of silica-gel and Sephadex LH-20 gel column chromatographies: 400 MHz <sup>1</sup>H NMR ( $CDCl_3$ – $CD_3OD$ , 2:1 v/v; 7.26 ppm-singlet of  $CHCl_3$  as an internal standard)  $\delta$  1.14 [coherent peak, 60H,  $(CH_2)_{30}$ ], 2.45 (s, 6H, 2 $\times$ *sn*-2-CH); IR (KBr) 1735 (C=O), 1240 and 1095 (P–O–C)  $cm^{-1}$ . Table 1 lists physical constants of **2**–**7**.

**Biological Assays.** (i) **Platelet Aggregation.** A 9:1 volumetric mixture of male New Zealand white rabbit blood and 3.8% trisodium citrate was centrifuged at 1100 rpm for



10 min at room temperature. The upper layer, platelet-rich plasma (PRP), was collected. Then, the residual blood sample was centrifuged at 3000 rpm for 10 min at 4°C to provide platelet-poor plasma (PPP). Platelet count in PRP was measured by Systemax Platelet Counter PL-100, and adjusted to  $5 \times 10^8$ /ml by appropriate dilution with PPP. Platelet aggregation was initiated by adding 30  $\mu$ l of **7** in a 0.025 M (1 M = 1 mol dm<sup>-3</sup>) tris-HCl buffer (pH 7) containing 0.13 M NaCl into 270  $\mu$ l of PRP. The assay was conducted according to the turbidimetric procedure of Born and Cross.<sup>13</sup> In the experiment to know the effect of *rac*-3-[*N*-(octadecyl)carbamoyloxy]-2-methoxypropyl 2-(3-thiazolino)-ethyl phosphate (CV-3988)<sup>12</sup> on test compound-induced platelet aggregation, PRP was pretreated with 100  $\mu$ M of CV-3988 for 3 min. Typical results are displayed in Fig. 1.

(ii) **Antihypertension.** Wister male rats (weight, 254–276 g) were anesthetized with sodium pentobarbital (60 mg/kg, i.p.). Appropriate amounts of **7** dissolved in saline or

PAF in 0.05 M tris-HCl–0.15 M NaCl buffer (pH 7) was then injected intravenously, and the blood pressure in jugular vein was measured for 15 min by the use of a pressure-transducer, a Nippon Hatsuden model RMP-6018. The results are shown in Fig. 2.

## Results and Discussion

It was found that dimerization of PAF into **7** resulted in modifying the platelet aggregation and antihypertension activities in a unique manner; e.g., although the maximum levels of both activities of **7** were 1/50–1/100-fold of those of PAF (Fig. 1a and 1b; Fig. 2) and appeared to be dose-dependent in the examined range ( $10^{-1}$ –1  $\mu$ M for platelet aggregation and 10–100  $\mu$ g/kg of rat for antihypertension), the antihypertension effect was exhibited with time lag of

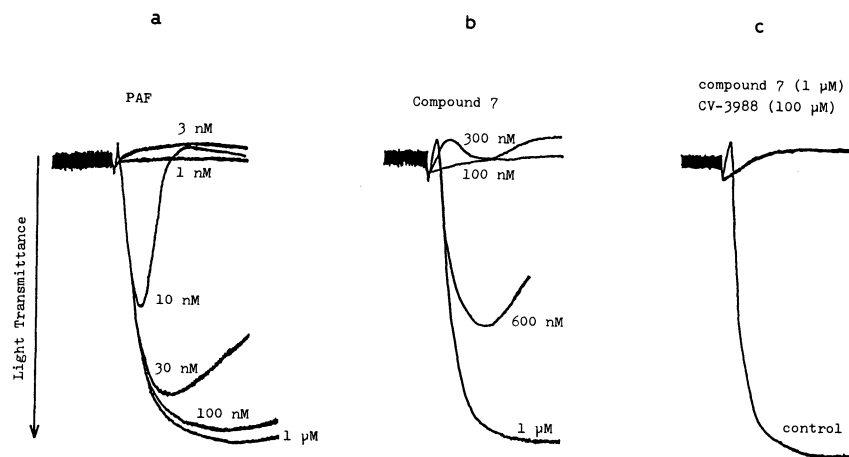


Fig. 1. Typical patterns of PAF- and compound **7**-induced rabbit platelet aggregation (a and b), and inhibitory effect (c) of CV-3988 on the compound **7**-induced aggregation.

Table 1. Physicochemical Results<sup>a)</sup>

Compound	Purification	Yield %	Mp/°C	$[\alpha]_D^{25}$	Silica-gel TLC, $R_f$	C and H analysis FAB mass spectrum
<b>2</b>	Silica-gel column <sup>b)</sup>	73	69–72	–6.9° (c 1.3, chloroform)	0.37 <sup>h)</sup>	$m/z$ 711 (M+1) (Calcd for C <sub>44</sub> H <sub>86</sub> O <sub>6</sub> , 710).
<b>3</b>	Recrystal. (chloroform)	95	112–117	–2.8° (c 1.5, chloroform)	0.35 <sup>i)</sup>	Calcd for C <sub>38</sub> H <sub>78</sub> O <sub>6</sub> : C, 72.33; H, 12.46%. Found: C, 71.90; H, 12.53%.
<b>4</b>	Silica-gel column <sup>c)</sup>	77	g)	+2.6° (c 1.25, chloroform)	0.39 <sup>j)</sup>	$m/z$ 1115 (M+1) (Calcd for C <sub>76</sub> H <sub>106</sub> O <sub>6</sub> , 1114).
<b>5</b>	Silica-gel column <sup>d)</sup>	92	g)	+11.7° (c 1.7, chloroform)	0.61 <sup>j)</sup>	$m/z$ 1199 (M+1) (Calcd for C <sub>80</sub> H <sub>110</sub> O <sub>8</sub> , 1198).
<b>6</b>	Sephadex LH-20 gel column <sup>d)</sup>	85	77–79	–3.5° (c 3.5, chloroform)	0.23 <sup>k)</sup>	$m/z$ 715 (M+1) (Calcd for C <sub>42</sub> H <sub>82</sub> O <sub>8</sub> , 714).
<b>7</b>	Silica-gel column, <sup>e)</sup> then Sephadex LH-20	8	241–244	–3° (c 0.3, chloroform– methanol, 2:1 v/v)	0.40 <sup>l)</sup>	$m/z$ 1045 (M+1) (Calcd for C <sub>52</sub> H <sub>106</sub> O <sub>14</sub> N <sub>2</sub> P <sub>2</sub> , 1044)

a) Silica-gel column support, Merck 7734, 70–230 mesh; Silica gel TLC, precoated sheet, Merck Art. 5735; matrix used in the FAB mass spectra, 3-mercapto-1,2-propanediol–glycerol, 1:1 v/v. b) Hexane–ethyl acetate–triethylamine in gradually changing from 70:4:1 to 40:4:1 v/v. c) Hexane–ethyl acetate in gradually changing from 13:1 to 3:1 v/v. d) Chloroform–methanol, 2:1 v/v. e) Chloroform–acetone–methanol–conc'd ammonia in gradually changing from 8:6:6:1 to 8:6:6:3 v/v. f) Hexane–ethyl acetate, 7:1 v/v. g) A compound to melt at room temperature. h) Hexane–ethyl acetate, 5:1 v/v. i) Chloroform–methanol, 8:1 v/v. j) Hexane–ethyl acetate, 3:1 v/v. k) Chloroform–ethyl acetate, 1:1 v/v. l) Chloroform–acetone–methanol–conc'd ammonia, 4:2:6:3 v/v.

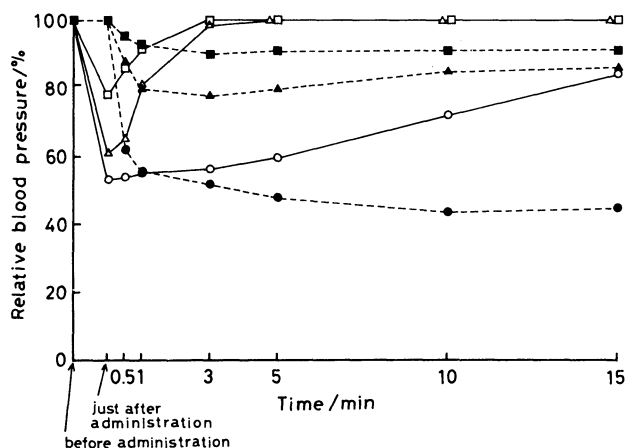


Fig. 2. Time course of antihypertension effects of compound **7**, 100 µg/kg (●), 30 µg/kg (▲), 10 µg/kg (■), PAF, 1.0 µg/kg (○), 0.3 µg/kg (△), and 0.1 µg/kg (□) in rats. Each point represents the mean of three animals.

1–3 min and seemed to persist for a period much longer than PAF (Fig. 2). By contrast, the effect of PAF was transient and became inactive rapidly within a few minutes after administration. The integrated biological activity over the effective time, though examined only qualitatively, may thus be comparable with or surpass that of PAF. A mechanism of the PAF-effect has not been understood well.<sup>12,14–16</sup> The compound **7**, however, was seemed to act on platelets in a manner similar to PAF because (i) **7** resembled with PAF in aggregation pattern and (ii) the activity of **7** was inhibited competitively by an antagonist of PAF, CV-3988<sup>14</sup> (typical aggregations: Fig. 1a–c). The decreased magnitudes in the biological activities of **7** may be attributed to conformational or steric effect of the 1,32-dotriacontanediyl residue and/or the increased polarity due to two phosphorylcholine residues. Such a structural modulation would result in slow metabo-

lism leading to long duration of the activities, and the time lag as well, which might be ascribed to slow accommodation of **7** to PAF-receptors on platelets.

#### References

- 1) C. A. Demopoulos, R. N. Pinckard, and D. J. Hanahan, *J. Biol. Chem.*, **254**, 9355 (1979).
- 2) J. Benveniste, M. Tence, P. Varenne, J. Bidault, C. Boullet, and J. Polonsky, *C. R. Acad. Sci. Paris, Ser. D*, **289**, 1037 (1979).
- 3) R. N. Pinckard, L. M. McManus, and D. J. Hanahan, "Adv. Inflam. Res.," ed by G. Weissmann, Raven Press, New York (1982), Vol. 4, pp. 147–180.
- 4) M. Shiraiwa, K. Fujita, H. Yoshiwara, S. Kobayashi, and M. Ohno, *Yuki Gosei Kagaku Kyokai Shi*, **45**, 369 (1987).
- 5) D. J. Hanahan, P. G. Munder, K. Satouchi, L. M. McManus, and R. N. Pinckard, *Biochem. Biophys. Res. Commun.*, **99**, 183 (1981).
- 6) G. Hirth, H. Saroka, W. Bannwarth, and R. Barner, *Helv. Chim. Acta*, **66**, 1210 (1983).
- 7) P. Harvary, J. M. Cassal, G. Hirth, R. Baener, and H. R. Baumgartner in INSERM Symposium 23, Platelet Activating Factor and Structurally Related Ether Lipids, ed by J. Benveniste and B. Arnoux, Elsevier, Amsterdam, 1983, p. 57.
- 8) J. J. Godfroid, F. Heymans, E. Michel, C. Redeuilh, E. Steiner, and J. Benveniste, *FEBS Lett.*, **116**, 161 (1980).
- 9) K. Satouchi, R. N. Pinckard, L. M. McManus, and D. J. Hanahan, *J. Biol. Chem.*, **256**, 4425 (1981).
- 10) M. Ohno, K. Fujita, H. Nakai, S. Kobayashi, K. Inoue, and S. Inoue, *Chem. Pharm. Bull.*, **33**, 572 (1982).
- 11) A. Pluckthun and E. A. Dennis, *Biochemistry*, **21**, 1743 (1982).
- 12) Z. Terashita, Y. Imura, and K. Nishikawa, *Biochem. Pharmacol.*, **34**, 1491 (1985).
- 13) G. V. R. Born and M. J. Cross, *J. Physiol.*, **168**, 178 (1963).
- 14) S. Hwang, C. Lee, M. J. Cheah, and T. Y. Chan, *Biochemistry*, **22**, 4756 (1983).
- 15) P. Inarrea, J. G. Gambroner, M. Neito, and M. S. Crespo, *Eur. J. Pharmacol.*, **105**, 309 (1984).
- 16) F. H. Valone, E. Coles, V. R. Reinhold, and E. J. Goetzl, *J. Immunol.*, **129**, 1637 (1982).